

# PREOMICS

## iST Fractionation Add-on

### Washed peptides



### Introduction

Sample preparation is one of the essential steps of bottom-up proteomics. The PreOmics iST sample preparation kit is designed to assist researchers achieving best results with few sample preparation steps and little hands-on time. For sample-specific protocols and optimization visit [www.preomics.com/downloads](http://www.preomics.com/downloads) or contact [info@preomics.com](mailto:info@preomics.com).

### Kit Contents

The kit contains everything to perform peptide fractionation into three fractions.

Component	Cap	Quantity			Buffer Properties				Description	Storage
		8rxn	12rxn	96rxn	Organic	Acidic	Basic	Volatile		
FRACTION-1	●	1x 2 mL	2x 2 mL	12x 2 mL	●		●	●	Fractionates the peptides from the cartridge, fraction 1.	RT
FRACTION-2	●	1x 2 mL	2x 2 mL	12x 2 mL	●		●	●	Fractionates the peptides from the cartridge, fraction 2.	RT
FRACTION-3	●	1x 2 mL	2x 2 mL	12x 2 mL	●		●	●	Fractionates the peptides from the cartridge, fraction 3.	RT

### Pre-Requisites

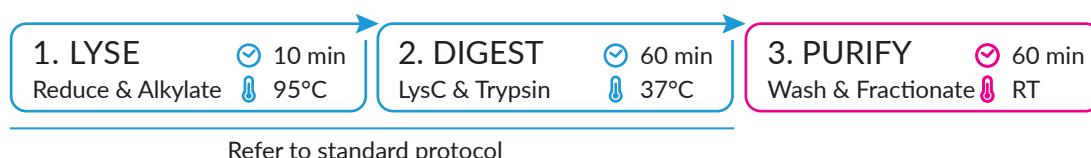
Common lab equipment is required for the sample preparation.

#### Equipment

#### Quantity and Description

PIPETTE	Careful sample handling and pipetting reduces contaminations and improves quantification.
SAMPLE	Washed peptides bound to the cartridge.
CENTRIFUGE	1.5/2.0 mL reaction tube centrifuges are required for elution.
VACUUM EVAPORATOR	Vacuum manifolds evaporate volatile buffers from the eluate before LC-MS.
ULTRASONIC BATH	Optional: can be used to resuspend peptides.

### Procedure



## Method

### 1. LYSE

1.1. Refer to standard protocol

### 2. DIGEST

2.1. Refer to standard protocol

### 3. PURIFY

3.1. to 3.4. Refer to standard protocol

3.5. Add 200  $\mu$ L **FRACTION-1** ● to **CARTRIDGE**, spin in CENTRIFUGE (1,000 rcf, 1 min).

Keep flow-through in **COLLECTION** tube 1. Transfer **CARTRIDGE** to a new **COLLECTION** tube.

3.6. Add 200  $\mu$ L **FRACTION-2** ● to **CARTRIDGE**, spin in CENTRIFUGE (1,000 rcf, 1 min).

Keep flow-through in **COLLECTION** tube 2. Transfer **CARTRIDGE** to a new **COLLECTION** tube.

3.7. Add 200  $\mu$ L **FRACTION-3** ● to **CARTRIDGE**, spin in CENTRIFUGE (1,000 rcf, 1 min).

Keep flow-through in **COLLECTION** tube 3.

3.8. Discard **CARTRIDGE** and place **COLLECTION** tubes 1-3 in a vacuum evaporator (45°C; until completely dry).

3.9. Add **LC-LOAD** ○ to **COLLECTION** tubes 1-3. Aim for 1 g/L concentration (e.g. 100  $\mu$ L to 100  $\mu$ g protein starting material).

**\*NOTE 1\***

3.10. Sonicate **COLLECTION** tubes 1-3 in an **ULTRASONIC BATH** (5 min) or shake (RT; 500 rpm; 5 min). **\*SP\***

**\*Note 1\*:** Because of the nature of the buffers, we recommend using BCA assay rather than measuring the peptide concentration with absorption (A260 or A280).

**\*SP\* - Storage Point:** At this point, close the peptide containing tube or **CARTRIDGE** using silicon lid.  
Peptides can be frozen at -20°C. Storage of peptides should not exceed two weeks at -20°C.  
For extended storage, finish the protocol and store at -80°C.

## Data analysis

Consider the fixed modifications as referred to in the corresponding kit protocol

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